Supplemental Figures.



Supplementary Figure 1. Polyacrylamide matrices that are (**a**) low polymer content and soft; (**b**) high polymer content and stiff; and (**c**) high polymer content, but reduced stiffness due to incorporation of fluorescein o-methacrylate, a chain-terminating monomer. (**d**) Bulk shear rheology on polyacrylamide hydrogels polymerized with fluorescein o-methacrylate monomers demonstrate that for 100 μ g/mL of fluorescein methacrylate, the stiffness of polyacrylamide matrices are reduced by five-fold compared to non-fluorescent hydrogels. The modified hydrogels retain their mechanical modulus properties over a large deformation range.



Supplementary Figure 2. Characterization of MSG fluorescence through co-polymerization with fluorescein methacrylate (fMA) monomers. Hydrogel fluorescent intensity depends on (**a**) MSG modulus, and (**b**) fMA concentration, as more polymerized fluorescent monomer results in higher fluorescent intensity. (**c**) Surface functionalization and sterilization steps further reduce the fluorescent intensity after the hydrogel is washed thoroughly. Data reported as mean \pm standard deviation. These results indicate that fluorescein methacrylate is incorporated stably into the polymer backbone, allowing persistent labelling during cell culture experiments.



Supplementary Figure 3. Confirmation of cell viability in polyacrylamide hydrogels modified with fluorescein methacrylate monomers. HS-5 human bone marrow fibroblast cells cultured on polyacrylamide hydrogel formulations remained viable after 24 hours independent of (a) hydrogel modulus, and (b) fluorescent methacrylate concentration within the hydrogel. Data reported as mean \pm standard deviation.



Supplementary Figure 4. Production and characterization of polydisperse hydrogel MSGs. (**a**) A two-phase oil/water immiscible system was used for the facile fabrication of hydrogel microspheres. Polyacrylamide pre-polymer with fluorescein methacrylate monomers were mechanically dispersed in an immiscible kerosene phase and allowed to polymerize under gentle stirring. (**b**) Size distribution of recovered microspheres indicate normal distribution of sizes with a mean diameter of ~50 µm.



Supplementary Figure 5. Size separation of polydisperse hydrogel MSGs by sequential centrifugation. (a) Sequential centrifugation steps can be used to separate smaller hydrogel microspheres (scale bar = $50 \mu m$), (b) as characterized by decreasing mean diameters, n = 100.



Supplementary Figure 6. Production of monodispersed hydrogel MSGs via microfluidic droplet generation. (a) A microfluidic droplet generator system consisting of a pulled circular micropipette inserted into a square glass tube was used to synthesize hydrogel microspheres of relatively uniform size. Droplets of polyacrylamide pre-polymer with fluorescein methacrylate monomers were generated in kerosene. A catalyst was added downstream of the droplet generation site, and hydrogel microspheres were allowed to polymerize. (b) The process enables the rapid production of a large number of uniform hydrogel microspheres (scale bar = $200 \mu m$).



Supplementary Figure 7. Volumetric swelling ratio of hydrogel MSGs in oil and aqueous phases. The soft hydrogels developed here swell significantly in aqueous medium, making it challenging to fabricate very small MSGs of uniform size. Data reported as mean \pm standard deviation, n > 20, * indicates p < 0.05 (one-way ANOVA with Tukey post-hoc pairwise comparisons).



Supplementary Figure 8. Shear rheometry characterization of bulk, fluorescently-labelled polyacrylamide hydrogels. These bulk measurements demonstrate linear elastic material properties with negligible loss modulus (**a**) over large strain range (data repeated in main manuscript Fig. 2a) and (**b**) for load frequencies less than 1 Hz. For frequencies greater than 1 Hz, the reduced shear modulus is likely due to slippage of the hydrogel on the rheometer plates. However, frequencies greater than 1 Hz are unlikely to be relevant to the current application of MSGs in multicellular spheroids.



Supplementary Figure 9. Measurement of Poisson's ratio of the fluorescently-labelled polyacrylamide hydrogel formulations used in this work. (a) Polyacrylamide hydrogel "strings" were fabricated within glass capillaries (internal diameter of 1.3-1.6 mm) that had been pretreated to be hydrophobic. Following gelation, polyacrylamide hydrogel strings were released from the glass capillary and swelled for 24 hours before (b) stretching axially (scale bar = 1 mm) under a fluorescent dissecting microscope. The deformations in the transverse and axial directions were measured to compute the Poisson's ratio which (c) remains constant for strains up to 120% (data repeated in main manuscript Fig. 2b).



Supplementary Figure 10. Measurement of MSG mechanical properties by application of osmotic pressure. An aqueous solution of long-chain dextran (500 kDa) was used to exert an osmotic pressure on polyacrylamide hydrogels. The dextran polymer chains are too large to enter the polyacrylamide pores, and are therefore excluded from the hydrogel. The osmotic pressure differential forces water out of the MSG, which deforms in proportion to the MSG mechanical compliance. (a) A schematic representation and (b) fluorescent microscope images (scale bar =50 μ m) depicting hydrogel contraction when exposed to 100 mg/mL of dextran solution. (c) MSG sizes remain constant after 3 hours in the dextran solution, confirming that dextran chains are excluded from the polymer matrix (n = 19). The system was calibrated against osmotic pressure-induced deformation of a bulk disk-shaped polyacrylamide hydrogel sample (diameter = 13 mm) for which the shear modulus was established using conventional shear rheometry. (d) A finite element simulation was developed to determine the effective osmotic pressure generated by a 100 mg/mL solution of dextran. The parametric sweep of external pressures on samples was used to determine that 100 mg/mL of dextran exerts 67 Pa pressure on the hydrogel surface. Next, this osmotic pressure value was applied to (e) a parametric sweep of shear modulus in the isotropic compression of a spherical MSG. (f) Osmotic pressure measurements on MSGs indicates that collagen coating does not significantly alter mechanical rigidity of the MSG (n =24, p = 0.782). (g) No significant differences were found between coated MSGs (control) and MSGs that had been removed from spheroids after two days of culture by detergent-based extraction (released), demonstrating that MSG properties remain constant even after embedding within the tissue of interest (n = 16-19, p = 0.837). All data reported as mean \pm standard deviation. NS indicates no significant differences (one-way ANOVA with Tukey post-hoc pairwise comparisons).



Supplementary Figure 11. Finite element model to simulate multiaxial MSG deformation. (a) Schematic of the two-dimensional axisymmetric model along with strain conditions applied throughout the bead domain. (b) Representative image of a partially revolved axisymmetric MSG bead deforming under -0.33 axial strain and -0.5 radial strain domain conditions. Corresponding (c) axial and (d) radial stresses are confirmed to be uniform throughout the MSG, consistent with the assumption of viscous flow in the surrounding tissue.



Supplementary Figure 12. Compressibility of the hydrogel is an important parameter in generating unique solutions for axial and radial stresses based on measured MSG deformation. Finite element simulations relating axial (z) and radial (r) stresses with axial and radial strains for (a) perfectly compressible (v = 0), (b) actual (v = 0.3) and (c) incompressible (v = 0.499) materials. For perfectly compressible materials, strains in the axial and radial directions are only weakly coupled to radial and axial stresses respectively. On the other end of the spectrum, as the material approaches (c) incompressibility (v = 0.499), microsphere deformations cannot be resolved into unique combinations of axial and radial stress. Hence, the use of compressible materials enables the measurement of both isotropic and anisotropic stress components in the system.



Supplementary Figure 13. Characterization of MCS structure 48 hours after formation. (a) Type I collagen fluorescent immunostaining on sectioned MCS confirms that HS-5 fibroblasts secrete Type I collagen over 2 days of culture (scale bar = $250 \mu m$). Negative control performed without primary antibody confirms that the signal detected is not a result of non-specific binding. (b, c) Second harmonic imaging of collagen shows no spatial variation in ECM organization within spheroid sections. (b) Second harmonic imaging of mature collagen on sectioned spheroids indicates no spatial variations (scale bar = $100 \,\mu$ m). (c) Quantification of fluorescence intensity along a line segment and normalized to quantified area shows no variation in mature collagen content within MCS. (d, e) The core of spheroid cultures does not exhibit hypoxia as indicated by immunohistochemical analysis for carbonic anhydrase 9 (CA9), a marker of hypoxia. (d) A representative immunohistochemical section is shown (scale bar = 50 μ m). (e) Immunohistochemical analysis shows similar abundance of CA9 in cells located in the periphery and the core of the spheroid, indicating an absence of an oxygen gradient. Data reported as mean \pm standard deviation, n = 13, * indicates p < 0.001 (one-way ANOVA with Tukey post-hoc pairwise comparisons). (f) Characterization of cell elongation in H&E stained spheroid sections. Data reported as mean \pm standard deviation, n = 30 over 3 spheroids, * indicates p < 0.001 (oneway ANOVA with Tukey post-hoc pairwise comparisons).



x-z plane

Supplementary Figure 14. Reconstructed confocal images of MSGs embedded at the periphery of the MSG confirms symmetric deformaton of the MSGs within spheroid cultures. (a) Schematic representation of the MSG location within the spheroids, and (b) reconstructed confocal images of MSGs close to the surface show deformations as expected based on spherical symmetry (scale bar = $25 \mu m$), (c) The 'pancake'-like morphologies adopted demonstrate two main axes of uniform deformation (radial and circumferential), arising from compressive radial stress and tensional circumferential stress.



Supplementary Figure 15. Measurement of circumferential and radial microsphere strains indicate some spatial pattern in microsphere orientation within blebbistatin treated MCS in the first 24 hours. This preferential orientation is lost by day 2 in blebbistatin treated MCS cultures. Data reported as mean \pm standard deviation for measurement error.



Supplementary Figure 16. Comparison of MSG errors associated with uncertainties in MSG modulus (accuracy) and strain measurement error (precision) in the (**a**) radial and (**b**) circumferential directions at day 2 of culture. Red data points represent tensional stress measurements, blue data points represent compressional stress measurements, and black data points represent stress measurements close to zero (-10 Pa to +10 Pa). Insets depict closer view of measured tensional stresses. Accuracy errors correspond to errors of 6% in stress readings, while precision errors were generated based on Monte Carlo simulations of error assuming a Gaussian normal distribution of values for repeated measurements of radial and circumferential bead dimensions. Both errors are combined and reported in the main manuscript figures (Fig. 4).



Supplementary Figure 17. Representative bar graphs of Monte Carlo estimates in stress measurement uncertainties arising from errors in measurement of MSG deformation. Repeated measurement of MSG dimensions was used to estimate the error in analysis of MSG size along the axial and radial axes. Assuming a Gaussian normal distribution of measurements in both the radial and circumferential axes for each data point, 10,000 randomly generated deformation values were converted to stresses through the non-linear interpolation function described in Fig. 2e, f. (a, b) Representative datasets from (a) axial and (b) radial stress Monte Carlo statistical distributions for a single axial compression-radial tension MSG data point (-6.50 Pa in the axial direction; +40.46 Pa in the radial direction). Mean stress values (dashed line) and their respective 95% confidence intervals (green section) are obtained empirically from the randomly generated dataset around each point. Similar curves were generated for every datapoint analyzed, and the 95% confidence intervals for each point are plotted as estimates of error in Supplementary Figure 16. These errors are then combined with errors in systemic accuracy to determine the total measurement error, values reported in Fig. 4, and in supplemental tables.



Supplementary Figure 18. Finite element model to simulate cell-generated mechanical stresses within MCS cultures driven by differences in cell proliferation. Finite element simulations show internal stress profiles generated with (1) iso-volumetric growth in the edge, intermediate zone, and core regions; (2) a non-proliferative edge with the intermediate zone and core growing at the same rate; (3) a non-proliferative edge with the core growing faster than the intermediate zone; and (4) a non-proliferative edge with the core growing slower than the intermediate zone. The stress profiles indicate that a non-proliferating edge is required to obtain a shell of tension in the circumferential direction around the outer layer of the MCS, as the static edge confines the growing internal layers of the spheroid, much like the walls of an inflating balloon. Mismatches in growth between the intermediate zone and core generate different stress profiles, where a peak in compression can only be obtained when the core is growing at a slower rate than the intermediate zone.



Supplementary Figure 19. The intermediate zone of the spheroid does not exhibit maximal cell proliferate rate by immunohistochemical analysis of Ki67, a marker of cell proliferation. (**a**) A representative immunohistochemical section is shown (scale bar = 50 µm). (**b**) Immunohistochemical analysis shows significantly higher percentage of cells positive for nuclear Ki67 in the periphery compared to the core of the spheroid, suggesting an increased rate of proliferation towards the edge of the spheroid (scale bar = 50 µm). Data reported as mean \pm standard deviation, n = 11, * indicates p < 0.05 when compared to other spatial locations (one-way ANOVA with Tukey post-hoc pairwise comparisons). (**c**) Volumetric expansion after one cycle of cell division calculated based on nuclei packing and Ki67 staining results. Data reported as mean \pm standard deviation, n = 9, * indicates p < 0.05 (one-way ANOVA with Tukey post-hoc pairwise comparisons).



Supplementary Figure 20. Confirmation of blebbistatin activity through quantification of spheroid size. Compaction is significantly reduced in MCS when actomyosin contractility is inhibited with blebbistatin (day 1), but is regained when the drug is washed out (day 2). Data reported as mean \pm standard deviation, n = 11, * indicates p < 0.01 (Student's t-test).

Supplem	entary Table 1.	Radial and circumferential strains for control data set.							
				Radial strain	Circumferential				
	Distance to	Padial strain	Circumferential	standard	strain standard				
Day 0	253,271667	-0.0173117	-0.0173117	0.00750415	0.00750415				
Dayo	125.839	0.03100127	0.03100127	0.01065199	0.01065199				
	22.1433333	-0.0185852	-0.0185852	0.0095284	0.0095284				
	80.5896667	0.0153115	0.0153115	0.00577389	0.00577389				
	250.139	-0.0434611	-0.0434611	0.0090087	0.0090087				
	72.9496667	-0.0146369	-0.0146369	0.01624392	0.01624392				
	78.2076667	-0.023329	-0.023329	0.01774364	0.01774364				
	533.014333	-0.0119751	-0.0119751	0.01218096	0.01218096				
_	79.9646667	-0.0024966	-0.0024966	0.00878492	0.00878492				
Day 1	264.881667	-0.0756392	-0.3650069	0.01475975	0.02080695				
	201.624667	-0.4435282	-0.2124585	0.01250441	0.0142695				
	102.540667	-0.1189901	-0.4600942	0.00701969	0.01079038				
	192.022333	-0.2914491	-0.4571600	0.01000001	0.00860798				
	1/2 887333	-0.4044002	-0.1407770	0.0301293	0.00809539				
	215 912333	-0.0337602	-0.3270834	0.001718	0.01150695				
	112.064	-0.1931875	-0.3890554	0.0206755	0.00389932				
	163.003667	-0.1452642	-0.4215483	0.01621957	0.0095584				
	146.594333	-0.177023	-0.4728582	0.02426695	0.00460169				
	140.748	-0.1308068	-0.4913635	0.02469393	0.01827656				
	142.519	-0.2168688	-0.3912013	0.01278379	0.02710338				
	134.803333	-0.3127343	-0.2784181	0.00849512	0.01562575				
	155.406	-0.5408411	-0.38305	0.00354685	0.02167709				
	18.788	-0.1664079	0.12768318	0.0128192	0.02533022				
	293.869333	-0.2016779	-0.2935707	0.0148574	0.01092408				
David	319.972	-0.3827587	-0.1639899	0.00870317	0.01494147				
Day 2	100.0200	-0.2580223	-0.4048553	0.02098765	0.0081292				
	195.156007	-0.3040303	-0.3072777	0.01295048	0.01200982				
	146 8565	-0.3639815	-0.4997769	0.02346986	0.00832615				
	70.0875	-0.2281152	-0.5271602	0.01903957	0.00670429				
	124.007	-0.2009152	-0.3465628	0.03298533	0.02770151				
	86.7036667	-0.3705685	-0.2628322	0.0162228	0.00719932				
	57.3656667	-0.4689408	-0.2761605	0.00528345	0.00596835				
	138.430333	-0.2861065	-0.3040828	0.00647076	0.02165226				
	40.2276667	-0.3510402	-0.1272162	0.04559503	0.01905851				
	10.9856667	-0.2148388	0.31090689	0.0158255	0.00763435				
	85.64/666/	-0.504119	-0.2404744	0.01875568	0.02455746				
	160 01 4222	-0.4022993	-0.2001020	0.00037447	0.02107350				
	29 206	-0.2307270	-0.1164766	0.02129403	0.01303668				
	282.744	-0.1935859	-0.2687895	0.01394602	0.00398711				
	288.575667	-0.1706106	-0.3447979	0.02672242	0.02790839				
	127.933667	-0.2815952	-0.4925533	0.00332189	0.0138056				
	109.956333	-0.1099068	-0.4578969	0.0217457	0.01095201				
	192.460667	-0.2794271	-0.4438034	0.02203337	0.01126994				
	148.156333	-0.4242247	-0.1161216	0.02569421	0.05658027				
	189.080333	-0.1426186	-0.4670056	0.0267774	0.01341946				
	62.473	-0.3967844	-0.2068374	0.01197061	0.01966358				
	151.81/00/	-0.1/2903/	-0.4050082	0.02108204	0.015/0056				
	100.749333	-0.3130337	-0.4700091	0.00907175	0.01340130				
	193.000007	-0.2303100	-0.3401399	0.01047075	0.00517769				
	160 980333	-0.0868809	-0.3623651	0.01908261	0.01223852				
	118.081667	-0,3606838	-0.0797439	0.02692123	0.02829732				
	133.631667	-0.3666899	-0.087223	0.01032526	0.02177059				
	18.941	-0.2678937	0.20217184	0.03220038	0.00766611				
	185.406	-0.4363358	-0.3340389	0.0146086	0.02669782				
	16.6946667	-0.2225128	0.24081644	0.01776937	0.02971592				
	229.6955	-0.1318971	-0.3409722	0.01519917	0.01715384				
	211.6955	-0.1148664	-0.3214346	0.01007977	0.01006305				

Supplementary Tabl	e 1. F	Radial an	d circum	ferential	strains	for	control	data se
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Suppi	ementur j	1 4010 20	itualai alla ell	cumerentiur	50105505 101	control data set	
				(-) Radial	(+) Radial	(-) Circumferential	(+) Circumferential
	Distance to	Radial	Circumferential	stress standard	stress standard	stress standard	stress standard
	edge (um)	stress (Pa)	stress (Pa)	deviation (Pa)	deviation (Pa)	deviation (Pa)	deviation (Pa)
Day 0	253.271667	-7.301111	-7.46598	2.673557	2.593557	3.083422	3.04337
	125.839	10.867287	10.744592	3.087121	2.934217	3.466865	3.361118
	22.1433333	-7.862515	-7.982986	3.387298	3.303862	3.817699	3.748761
	80.5896667	5.485827	5.396788	1.772409	1.794597	1.9167	1.947585
	250.139	-19.706897	-19.664205	4.412946	4.344097	4.804758	4.486302
	72 9496667	-6.208612	-6.239468	5 412413	5,225729	6 12386	5,721876
	78.2076667	-9.84552	-10.047527	6.671968	5.801549	7.252188	6.693407
	533.014333	-5.048624	-5.117344	3,832863	3.825252	4,514839	4.266862
	79 9646667	-1 076698	-1 120348	2 643903	2 435971	3 093488	2 799602
Day 1	264 881667	-261 75762	-334 43507	38 035727	52 732/01	47 45485	64 040733
Day I	204.001007	-201.75702	-344.45397	30.000727	53 707327	51 267018	50 /6076
	102 540667	521 06421	644 70621	27 256050	60 0027	65 560626	01 2001/2
	102.040007	-551.00451	-044.70031	S7.330039 60.937450	117 724707	111 500760	101.209140
	192.022333	-019.07040	-0/9.4/002	09.037439	117.734797	111.09Z70Z	125.020390
	122.090007	-327.09230	-200.0200	41.000437	00.300079	04.400740	47.000101
	142.887333	-210.22089	-127.97752	11.201104	23.744437	23.19474	17.009063
	215.912333	-179.82457	-245.99289	13.225495	23.783057	24.064874	32.007787
	112.064	-429.16753	-484.49093	27.561167	52.50293	51.578255	51.567937
	163.003667	-450.84812	-534.5096	33.832836	59.855218	56.470742	66.421246
	146.594333	-667.01141	-772.09647	47.534088	86.904745	83.335314	87.493245
	140.748	-663.59051	-796.52014	93.714852	132.293223	119.533007	152.127711
	142.519	-465.21639	-515.17424	75.8833	103.687071	94.450046	118.301684
	134.803333	-347.38045	-339.36633	29.590402	49.473281	45.958238	51.596852
	155.406	-1171.3065	-1112.9573	125.031734	193.401521	180.029051	194.988333
	18.788	-12.45476	20.202178	5.393404	5.971468	5.493387	7.784411
	293.869333	-263.00106	-283.75706	20.521845	35.563459	34.304281	37.504438
	319.972	-261.90548	-217.68424	19.624759	34.776661	32.433646	33.587842
Day 2	188.8285	-561.74726	-606.59078	74.345071	72.994292	74.597857	73.415059
	195.138667	-389.21268	-390.04614	51.764252	50.066928	53.518171	51.114409
	88.716	-692.93029	-775.17568	70.596834	69.651415	75.706811	73.828534
	146.8565	-1265.8421	-1324.417	176.092648	168,148629	175,724862	168.794673
	70.0875	-1058.3039	-1187.6877	129,106966	124,482019	136,100324	131,188514
	124.007	-346.58811	-383,7132	88,498538	75.11601	95.310407	80.613553
	86.7036667	-388.38562	-362,7006	47.290177	45.288914	43.406404	41.605168
	57.3656667	-569.80526	-517,59844	50.362442	50,991774	47.374011	47,727092
	138,430333	-363,20447	-367,80412	63,61905	59,24869	69.366227	64,504504
	40 2276667	-198 99379	-158 16559	53 440548	45 903549	42 860863	37 126817
	10 9856667	2 728722	51 664435	2 758705	2 605397	4 11539	4 110623
	85 6476667	-554 0227	-484 98441	104 268008	95 329613	99 417808	92 712085
	67 1053333	-/13 015227	-353 /7635	65 203254	50.020010	64 104445	58 100034
	160 01/1333	-200 06727	-177 51766	40 041258	37 310014	40 547420	37 110/182
	20 206	-200.30727	-104 83445	18 000161	17 607321	40.047429	16/00581
	29.200	224 40091	240 62026	24 951579	22 402270	22 772275	22 70/262
	202.144	214 27021	257 62040	79 699065	20.402079	20.770270	76 077075
	107 022667	-314.27931	-337.03049	120 00166	121 07024	152 029270	145 161452
	127.933007	-976.00019	-1001.7975	130.00100	72 024059	152.930279	140.101400
	109.900000	-510.55929	-024.00072	10.202414	10.024000	00.097009	01.037102
	192.460667	-736.68699	-793.52568	111.211723	105.124773	114.388202	109.361181
	148.156333	-249.70145	-189.79308	84.616506	69.884388	84.508178	69.713043
	189.080333	-587.96949	-699.56491	100.321726	94.225167	109.655728	104.467225
	62.473	-331.50159	-289.57881	52.55099	49.866248	51.475102	48.783613
	151.817667	-712.22459	-827.89834	135.846737	122.513602	152.064473	139.094313
	160.749333	-970.83425	-1032.3334	150.399661	140.79986	162.008984	151.48817
	193.553667	-387.43063	-415.59427	93.288523	81.157794	103.267326	90.685279
	198.174	-145.45285	-184.15277	24.225872	22.711096	22.790903	21.894121
	160.980333	-268.25797	-337.4293	41.56183	41.131887	47.77239	46.948338
	118.081667	-167.9813	-119.41575	38.930277	35.06345	35.279741	31.066598
	133.631667	-177.73477	-128.68484	29.776894	27.4939	28.42619	26.117421
	18.941	-21.819972	29.703175	9.180939	8.412283	4.856412	4.423935
	185.406	-664.25138	-633.86438	132.773095	119.351704	136.021535	120.612126
	16.6946667	-6.489743	40.463765	5.828272	5.302946	8.2697	7.276722
	229.6955	-272.29484	-322.97951	46.735394	43.332894	54.426264	50.12556
	211.6955	-230.56426	-278.4297	29.40033	28.336532	34.33804	33.707544

Supplementary Table 2. Radial and circumferential stresses for control data set.

				Radial strain	Circumferential
	Distance to		Circumferential	standard	strain standard
	edge (µm)	Radial strain	strain	deviation	deviation
Day 0	589.337333	-0.0036178	-0.0036178	0.01918289	0.01918289
	138.919	-0.0025079	-0.0025079	0.01554925	0.01554925
	256.589333	-0.021659	-0.021659	0.00523918	0.00523918
	470.431667	-0.0119578	-0.0119578	0.01186768	0.01186768
	418.662	-0.0001081	-0.0001081	0.02322107	0.02322107
	611.276	-0.0156326	-0.0156326	0.01735828	0.01735828
	198.466333	-0.0061013	-0.0061013	0.01056779	0.01056779
	19.0333333	0.00388331	0.00388331	0.00672608	0.00672608
	455.868	-0.0024674	-0.0024674	0.00427361	0.00427361
	295.665	0.0023396	0.0023396	0.01474246	0.01474246
Day 1	267.940333	-0.1459092	-0.0648114	0.02909371	0.03918094
	197.882333	-0.1331089	-0.3074382	0.01417008	0.00684715
	425.124	-0.167206	-0.17867	0.00350152	0.01024833
	218.567333	-0.2468586	-0.1183345	0.03166042	0.01812759
	158.978667	-0.084651	-0.1668022	0.02114879	0.01834155
	292.672	-0.0682101	-0.0880765	0.01041673	0.02072814
	111.107	-0.0684003	-0.2393817	0.01173793	0.01521672
	41.367	-0.2342072	-0.256783	0.02975877	0.01089485
	134.602333	-0.1505166	-0.2655213	0.01607685	0.01718629
	210.735	-0.0516536	-0.2303518	0.01723254	0.01605707
Day 2	233.825	-0.0610635	-0.0610635	0.03067846	0.03067846
	140.584333	-0.1354361	-0.2873418	0.00685044	0.01464093
	302.595333	-0.2047461	-0.2047461	0.01497081	0.01497081
	113.751667	-0.198659	-0.198659	0.01453806	0.01453806
	89.9246667	-0.1709642	-0.1709642	0.03567907	0.03567907
	253.883667	-0.1032205	-0.0301937	0.00805639	0.01423848
	118.448667	-0.2170043	-0.3249506	0.01525382	0.00647636
	52.81	-0.2141286	-0.1879506	0.01686422	0.02845634
	132.842	-0.3995903	-0.1918081	0.01627582	0.02307093
	186.460333	-0.2798243	-0.1945363	0.01028737	0.01708143

Supplementary Tab	le 3.	Radial	and	circumferentia	al strains	for	blebbi	statin	data	set.
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Supplementary radie 4. Kadiar and circumferentiar suesses for dieddistatin data set.									
				(-) Radial	(+) Radial	(-) Circumferential	(+) Circumferential		
	Distance to	Radial	Circumferential	stress standard	stress standard	stress standard	stress standard		
	edge (um)	stress (Pa)	stress (Pa)	deviation (Pa)	deviation (Pa)	deviation (Pa)	deviation (Pa)		
Day 0	589.337333	-1.617427	-1.53564	5.830154	5.317222	6.645397	5.760238		
	138.919	-1.14694	-1.117039	4.763156	4.326811	5.422539	4.694686		
	256.589333	-9.164581	-9.272991	2.197596	2.097677	2.399856	2.306334		
	470.431667	-5.06666	-5.135499	3.821231	3.854714	4.422385	4.421893		
	418.662	-0.221587	-0.193348	6.961076	6.206786	7.976663	6.779683		
	611.276	-6.552472	-6.61721	5.885485	5.487462	6.57209	6.176144		
	198.466333	-2.594393	-2.634016	3.349753	3.308872	3.843778	3.649243		
	19.0333333	1.383025	1.410119	1.824517	1.765593	2.202045	1.998116		
	455.868	-1.033383	-1.054274	1.313714	1.264916	1.541055	1.414794		
	295.665	0.774753	0.810081	4.262314	3.755217	4.913647	4.172076		
Day 1	267.940333	-57.369712	-46.043881	22.678407	20.125705	24.784701	22.112644		
	197.882333	-225.90097	-265.39404	27.329818	25.980629	29.09932	28.444571		
	425.124	-125.67903	-127.85501	14.864042	14.817605	16.789433	16.748918		
	218.567333	-127.10968	-105.62886	29.076092	27.043713	25.043378	23.088426		
	158.978667	-79.788391	-93.368335	18.454378	17.15671	20.705925	19.034733		
	292.672	-40.196489	-42.995932	10.514124	10.03789	13.476621	12.283223		
	111.107	-118.26448	-150.64298	20.241123	19.099798	24.998544	23.942445		
	41.367	-242.01016	-247.00051	43.390912	40.622641	39.940205	37.205777		
	134.602333	-189.86537	-213.88999	33.3809	31.325827	37.623777	35.295874		
	210.735	-102.94193	-135.80386	20.372866	18.422001	24.571439	22.64755		
Day 2	233.825	-29.461782	-29.561393	15.18428	13.624754	16.69375	14.927409		
	140.584333	-203.51195	-236.34333	30.316255	28.486122	36.13435	34.168667		
	302.595333	-167.4361	-167.49788	26.342316	25.133887	27.743775	26.001236		
	113.751667	-157.93236	-158.04869	24.702411	22.913386	25.954504	24.12351		
	89.9246667	-122.48514	-122.62976	40.798365	34.944301	43.514874	37.508318		
	253.883667	-32.312682	-22.770725	6.839203	6.282639	7.3087	7.102654		
	118.448667	-325.68243	-352.16013	37.930289	36.787076	38.195174	36.990667		
	52.81	-158.38726	-153.76607	35.760994	32.171951	39.741778	35.530578		
	132.842	-313.57802	-268.79854	55.496495	52.042927	54.0324	50.381177		
	186.460333	-209.06344	-192.22974	31.197826	30.365373	32.403349	31.062419		

Supplementary Table 4. Radia	l and	l circum	ferential	stresses	for	bleb	bistatin	data s	set.
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